

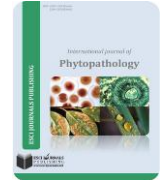


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SURVEY AND GENETIC DIVERSITY OF GRAPEVINE LEAFROLL ASSOCIATED VIRUS-2 IN ALGERIA

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ABSTRACT

Vineyards in western and center regions of Algeria were surveyed for the *Grapevine leafroll-associated virus 2* (GLRaV-2). Analyses by DAS-ELISA and Reverse Transcription Polymerase Chain Reaction (RT-PCR) reveal 15, 8% prevalence. The genetic diversity of the GLRaV-2 population was studied by phylogenetic analyses of the HSP70h gene region of seven samples sequenced in this study and other sequences downloaded from GenBank. Results reveal segregation of the GLRaV-2 population into six distinct groups. An estimation of the ratio of non-synonymous substitutions per non-synonymous site to synonymous substitutions per synonymous site indicated that HSP70h gene evolve under positive selection. Similarity plot constructed with representative sequence from each group confirmed previous results. All Algerian isolates belong to group PN. As far as we know, this is the first characterization of GLRaV-2 isolates from Algeria.

Keywords: GLRaV-2, RT-PCR, genetic diversity, HSP70h gene.

INTRODUCTION

The grapevine leafroll disease represents the most widespread disease of *Vitis vinifera* worldwide (Martelli, 2014). It may be due to several virus belonging to the family of *Closteroviridae* which were designated as *Grapevine Leafroll-associated Viruses* (GLRaVs) and represented by GLRaV-1, -2, -3, -4, and -7 which were recognized as species and recently (Martelli *et al.*, 2012) indicate that GLRaV-5, -6, -9, GLRaV-Pr, GLRaV-De, and GLRaV-Car are strains of GLRaV-4, considered before as distinct species. All these viruses belong to the family *Closteroviridae*, with GLRaV-2 belonging to the genus *Closterovirus*, GLRaV-7 belongs to a newly proposed genus *Velarivirus* and the other GLRaVs to the genus *Ampelovirus*. GLRaV-2 was described since 1984 and was associated with the grapevine leafroll disease (Gugerli *et al.*, 1984). Later, it was associated to graft-incompatibility (Greif *et al.*, 1995). The transmission vector for GLRaV-2 is unknown, although other members of the genus

Closterovirus were transmitted by aphids (Karasev, 2000). GLRaV-2 is known to be transmitted by grafting with infected material. However, some isolates of GLRaV-2 were mechanically transmitted to herbaceous hosts like *Nicotiana benthamiana* (Goszczynski *et al.*, 1996).

GLRaV-2 presents flexuous and filamentous particles of about 1600 nm length and its genome is a single strand, positive sense RNA with about 16500nt organized into nine ORF encoding at least 11 proteins (Zhu *et al.*, 1998). Up to now, complete genome sequence was obtained for nine isolates (JX559644.1, NC_007448.1, DQ286725.2, AY881628.1, KF220376.1, FJ436234.1, JQ771955.1, AF314061.1 and NC_004724.1).

Several variants of GLRaV-2 have been described and characterized. (Zhu *et al.*, 1998) and (Abou-Ghanem *et al.*, 1998) described the first two variants from the varieties Pinot Noir and Semillon. Two years later, a new variant denoted GLRaV-2-H4 was discovered by (Ghanem-Sabanadzovic *et al.*, 2000) on *Vitis rupestris* St. George. Further study reveals the presence of virus associated to GLRaV-2 isolated from cv Redglobe and associated to graft incompatibility (Rowhani *et al.*,

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2000). Up to now, six variants were reported, the variant 'Pinot Noir, 93/955, H4, BD, RG and PV20 (Zhu *et al.*, 1998, Meng *et al.*, 2005, Ghanem-Sabanadzovic *et al.*, 2000, Rowhani *et al.*, 2002, Angelini *et al.*, 2004, Beuve *et al.*, 2007, Bertazzon *et al.*, 2010).

MATERIAL AND METHODS

Virus Source: The field study and sample collection were conducted in autumn 2010 and 2012 on table grape and wine grape collected in western (Aïn Témouchent and Mascara) and central (Algiers, Tizi-Ouzou and Boumerdes) regions of Algeria. A total 584 samples were collected from individual vines from different varieties (30 varieties) including commercials (10 varieties) (445 samples) autochthonous vineyard (two varieties) (110 samples) and autochthonous Grapevine germplasm (18 varieties) (29 samples) of ITAF (Institute Technique de l'Arboriculture Fruitière et de la Vigne). Mature canes were randomly collected, one from each vine and stored at 4°C.

Virus detection by double antibody sandwich-enzyme-linked immunosorbent Assay (DAS-ELISA): All collected samples were tested by DAS-ELISA (Clark and Adams, 1977), for the presence of GLRaV-2 using specific commercial polyclonal antiserum (Agritest, Bari, Italy). The extracts were obtained by macerating phloem tissues in the PBS-buffer (V:V). Optical density was recorded at 405 nm using an automatic microplate reader (Multiskan Ascent, LabSystems, Waltham, MA, USA). Samples with Optical density readings exceeding or equal to three times that of the healthy samples were considered positive.

Virus detection by reverse transcription polymerase chain reaction (RT-PCR): All positive samples were analyzed by RT-PCR. Total nucleic acids were extracted from 0.2g of cortical scraping of dormant cutting cane, which was powdered using liquid nitrogen. The powder was homogenized in 1 ml of grinding buffer (4 M guanidine thiocyanate, 0.2 M sodium acetate, 1 M potassium acetate, 0.025 mM EDTA, 25% PVP-40) mixed with 100µl NLS 10% and denatured at 70°C. The solution was centrifuged and the supernatant was recovered. The Total Nucleic Acid was precipitated by Silica as described by (Svanella-Dumas *et al.*, 2000) with some modifications. Two step protocol was used for the reverse transcription (RT) and amplification (PCR) of target RNA. Reverse transcription was performed using 1 µl of Moloney murine leukaemia virus reverse transcriptase (M-MLV 200 units/ µl), 4 µl of 5× Fs M-

MLV buffer, 2 µl of DTT (0.1 M) and 0.5 µl of dNTPs (10 mM). The mixture was incubated at 39°C for 1h and at 70°C for 10 min. PCR was carried out with primer pair L2 F (5'-ATAATTCGGCGTACATCCCCACTT-3') and U2 R (5'-GCCCTCCGCGCAACTAATGACAG -3') which encompass 331pb located in the HSP70h gene. The DNA amplifications were carried out in 12,5 µl total reaction volume. 1,25 µl of cDNA were mixed with 11.25 µl of the amplification mixture, consisting of 10mM each dNTP, 20 µM each primer, 50 mM MgCl₂, 5 U/µl TaqDNA polymerase (Promega) and 10x Taq buffer. PCR cycling conditions, in a thermocycler, include denaturation of cDNA at 95°C for 5 min, followed by 35 cycles of 94°C/ 30s, 58°C/45s and 72°C/60 s. To end with, final elongation step was carried at 72°C during 7 min. The PCR products with positive and negative control used were provided from Institut Nationale de la Recherche Agronomique (INRAT) collection and PCR markers of 100pb were analyzed by electrophoresis on 1.5% agarose gel stained with ethidium bromide and visualized using an UV-transilluminator.

Sequencing and sequence analyses: Amplified products of 331pb from HSP70h gene region of GLRaV-2 of seven isolates were randomly chosen for sequencing, they were purified using the ExoSAP-IT purification kit. Direct sequencing was performed with the same primers used for RT-PCR on 3730xl DNA analyzer (Applied Biosystems) automated sequencer.

To investigate the different variants of GLRaV-2 population, multiple alignments of nucleotide sequences were performed using CLUSTAL W (Thompson *et al.*, 1994), respectively, with default settings from MEGA6 software (Tamura *et al.*, 2013) and comparison at the nucleotide level for HSP70h gene regions of Algerian isolates and representative sequences of the different phylogenetic groups described up to now was conducted (Table 2).

Phylogenetic analysis: Nucleotide sequences of the Algerian isolates and those downloaded from GenBank were aligned and evolutionary relationships among GLRaV-2 sequences inferred using the neighbor joining method (NJ) (Saitou and Nei, 1987) with 1,000 bootstrap replications. All these analyses were conducted in MEGA6 software with GLRaV-3 isolate BR5 (KF417599.1) used as an outgroup.

Estimation of selection pressure and recombination analysis: Gene and site-specific selection pressures over the entire alignment of data set for HSP70h were analyzed using the Datamonkey online (<http://www.datamonkey.org/>).

Table 1. Percent identity between Algerian isolates and isolates from each group.

Seq->	ALG7	ALG9	ALG19	ALG22	ALG53	ALG55	ALG93	PV20	OR1	GRSLaV	BD	PN	93/955	GLRaV-2-SG
ALG7	ID	98%	99%	97%	98%	99%	99%	74%	98%	71%	75%	99%	86%	85%
ALG9	98%	ID	99%	98%	99%	98%	99%	74%	99%	72%	74%	99%	86%	85%
ALG19	99%	99%	ID	99%	99%	99%	100%	75%	99%	72%	75%	100%	87%	86%
ALG22	97%	98%	99%	ID	98%	98%	99%	74%	99%	72%	74%	99%	86%	86%
ALG53	98%	99%	99%	98%	ID	98%	99%	74%	99%	72%	75%	99%	86%	86%
ALG55	99%	98%	99%	98%	98%	ID	99%	75%	99%	71%	75%	99%	87%	85%
ALG93	99%	99%	100%	99%	99%	99%	ID	74%	100%	72%	75%	100%	86%	86%
PV20	74%	74%	75%	74%	74%	75%	74%	ID	74%	71%	74%	74%	77%	74%
OR1	98%	99%	99%	99%	99%	99%	100%	74%	ID	72%	74%	100%	86%	86%
GRSLaV	71%	72%	72%	72%	72%	71%	72%	71%	72%	ID	81%	72%	71%	72%
BD	75%	74%	75%	74%	75%	75%	75%	74%	74%	81%	ID	75%	74%	77%
PN	99%	99%	100%	99%	99%	99%	100%	74%	100%	72%	75%	ID	86%	86%
93/955	86%	86%	87%	86%	86%	87%	86%	77%	86%	71%	74%	86%	ID	85%
GLRaV-2-SG	85%	85%	86%	86%	86%	85%	86%	74%	86%	72%	77%	86%	85%	ID

The ratio of nonsynonymous substitutions per nonsynonymous site (dN) to synonymous substitutions per synonymous site (dS), which is considered as an indicator of natural selection was calculated using two methods for detecting sites under positive selection: single-likelihood ancestor counting (SLAC), random-effects likelihood (REL). Depending on *dN/dS* values, the selection pressure was considered negative or purifying (*dN/dS* < 1), neutral (*dN/dS* = 1), or diversifying or positive (*dN/dS* > 1).

Recombinant events analysis of GLRaV-2 population was performed using the GARD algorithm (Kosakovsky *et al.*, 2006) in the remote server Datamonkey (Delpont *et al.*, 2010). A

similarity plot was constructed with Simplot software (downloaded from; <http://sray.med.som.jhmi.edu/SCSoftware/simplot/>) using the isolate OR1 from group PN as reference sequence with a multiple sequence alignment of full genome sequences provided from the different representative groups (H4, BD, RG, and 93/955) except for PV20 group constructed with MEGA6 (Tamura *et al.*, 2013). Currently, there is no full-length sequence for groups PV20.

RESULTS

Prevalence of GLRaV-2: DAS-ELISA test reveals that the GLRaV-2 presents prevalence of 15,8 % in Algeria. The peak of prevalence is observed in Gros noir des Beni Abbes with 29%. The Alicante

Bouchet presents 24,6%, King’s Rubi (22,2%), Dattier de Beyrouth (18,7%), Valensi (14,5%), Chaouch Blanc and Muscat d’Alexandrie (10%), Cinsault (7,5%), Carignan (9,7%), Cardinal (6,4%) and autochthonous collection (6,9%). The GLRaV-2 is absent in Merseguerra and Chasselas. The autochthones germplasm show to be free of GLRaV-2.

Sequencing and Sequence Analysis: The 331-nt sequence fragment of the HSP70h Gene obtained by RT-PCR was cloned and sequenced for seven isolates of GLRaV-2 representing the first sequenced isolates provided from Algeria. Nucleotide sequences obtained were submitted to GeneBank (Table 2).

Table 2. Range of HSP70h nucleotide sequence identities within and between groups of GLRaV-2.

Isolates	Average distance within groups						Average distance between groups					
	BD	PN	H4	RG	93/955	PV20	BD	PN	H4	RG	PV20	93/955
Distance	98%	99%	95%	96%	100%	100%	72-78%	70-6%	71-6%	69-78%	69-74%	71-86%

In order to access phylogenetic relationship between these isolates, comparison at nucleotide level was conducted with representative sequences from each group described by (Jarugula *et al.*, 2010). The Algerian isolates present 97% to 100% identities. Comparison with the different groups reveal 98% to 100% identity with the isolates OR1 and PN , 74% to 75% identity with the isolates PV20 and BD, 71-72% identity with GRSLaV, 86-87% identity with isolate 93/955, and 85- 86% identity with isolate GLRaV-2-SG.

The genetic distance, transformed to the percent identity between and within groups was calculated using MEGA6 software with default parameters reveals that the group PN presents 99% identity. The group H4 presents 95% identity, 96%, 98% and 100% identity for RG, BD and 93/955. The between percent identity reveal that the group PN presents 70-86% with all the other groups, 72-78% for group BD, 71-86%, 69-78%, 69-74% and 71-86% respectively for H4, RG, PV20 and 93/955 (Table 2).

Phylogenetic Analysis: Phylogenetic tree constructed using the neighbor joining method (NJ) implemented in MEGA6 software reveals that GLRaV-2 population cluster into six groups. Each of these Lineages were assigned to a reference isolate as described by previous study to maintain a standardized nomenclature of GLRaV-2 sequence variant groups. The groups PN and H4 represent the most important group of this population. The RG group contains isolates provided from the variety Red Glob obtained in California. The PV20, BD and 93/955 groups were less represented. Analyses reveal that all Algerians isolates collected from different regions, vineyards and varieties belong to the group PN. Further prospections in local varieties were needed to get a great understanding of genetic variation of GLRaV-2 in Algeria (Figure 1).

Similarity plot comparison of representative isolates from each group was conducted. Results reveal that the group PN is distant to the other groups and the groups H4 and 93/955 are more and less related, the same result was obtained for BD and RG groups which were found to be closely related (Figure 2). Currently, there are no full-length sequences from representatives isolates of groups PV20.

Selection Pressure: Identifying evolutionary pressure represent a great deal because past environments that exerted these pressures can be different from present ones and these pressures are not unidirectional but rather the result of complex networks (Moury and Simon, 2011).

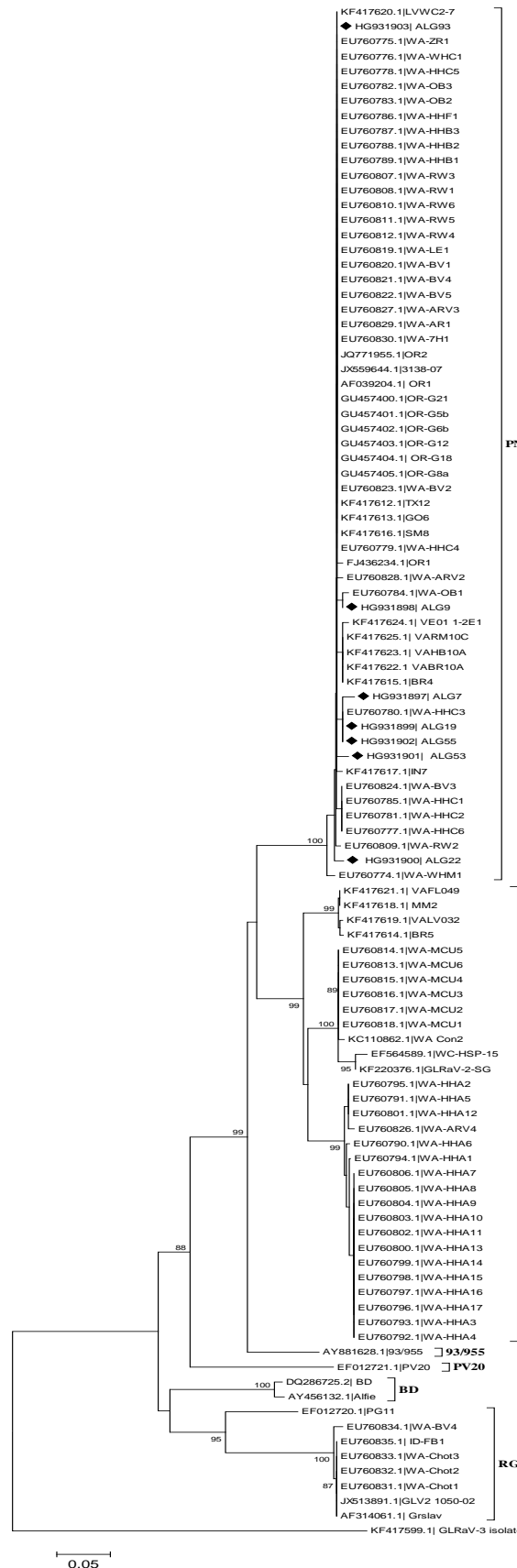


Figure 1. Phylogenetic tree of Grapevine Leafroll associated Virus 2 (GLRaV-2), based on HSP70h gene, constructed by the neighbor-joining method; The percentages of bootstrap support ($\geq 75\%$) from 1,000 replicates are shown at nodes.

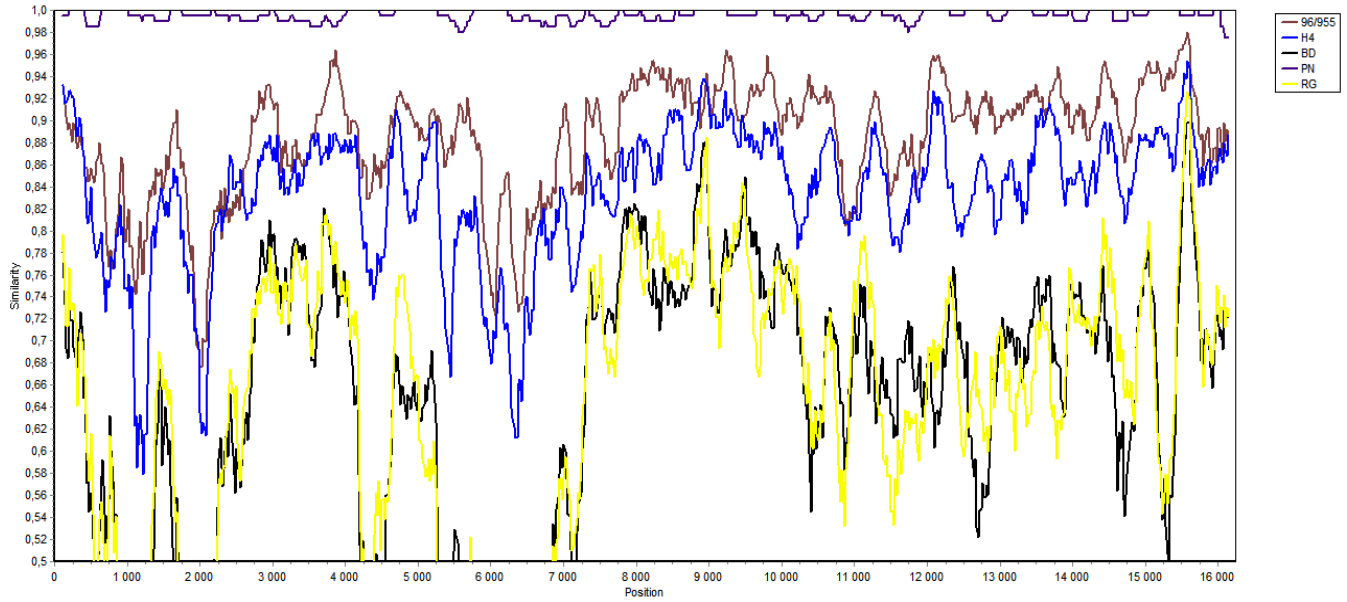


Figure 2. Similarity plot constructed from a multiple alignment of six full-length sequences representing five well-defined variant groups of GLRaV-2 using SimPlot 3.2.

In order to study selection pressure occurring in a GLRaV-2 population, an estimation of the ratio of nonsynonymous substitutions per nonsynonymous site (dN) to synonymous substitutions per synonymous site (dS) was performed. Indeed, dN/dS ratio indicates the magnitude of selective pressure on each codon. When the ratio $dN/dS > 1$ on a certain codon it indicates that the site evolves under positive selection and when $dN/dS < 1$ in another site it indicates that this site evolves under negative selection. The mean value of dN/dS ratio obtained with SLAC and REL methods is upper than unity suggesting the occurrence of positive selection. Indeed, SLAC reveals a mean ratio of $dN/dS > 1$ (1,81) with Codon sites under positive selection and non-negative selected codon

site were found. Approximately, same results were obtained by REL method with mean ratio $dN/dS = 3,71$ with 6 codons under positive selection and 6 codons under negative selection. Plotting of SLAC established by dN-dS for each codon, indicating the negative or positive selection, reveals that the number of codon sites under positive selection is more important compared to previous results obtained in this study (Figure 3). Detection of evidence of putative recombination events in the HSP70h sequences of the GLRaV-2 population was performed with the genetic algorithms for recombination detection (GARD) available on the Datamonkey webserver. GARD analysis reveals no recombinant events within a HSP70h gene in the GLRaV-2 population.

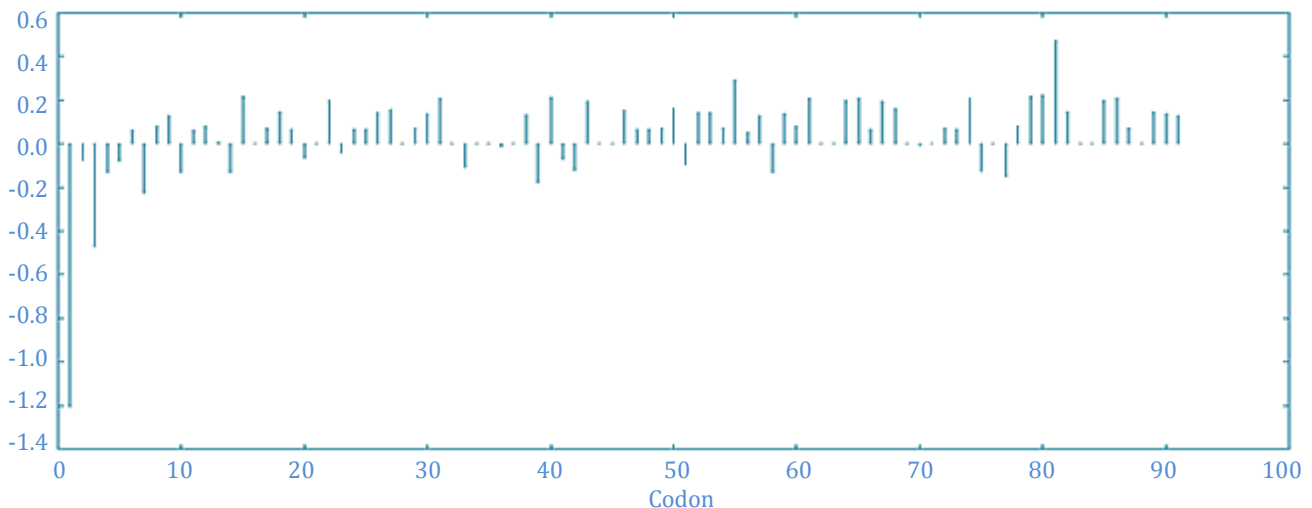


Figure 3. Plotting of single-likelihood ancestor counting (SLAC) based on 90 codon sites from HSP70h gene of GLRaV-2 sequences.

DISCUSSION

Survey and genetic diversity of GLRaV-2 collected from different vineyards in the center and western regions of Algeria was established. This study represents the first comprehensive work on the prevalence and genetic diversity conducted in major grape-growing region of Algeria including an autochthonous germplasm collection with first sequencing of Algerian isolates. Phylogenetic analyses with isolates provided from different regions was conducted to make an update of molecular variability of GLRaV-2.

Grapevine leafroll associated virus 2 is present in all prospected regions. This may be due to the use of infected root stock which in majority provide from North America. Indeed, the vector transmission of GLRaV-2 was not described up to now (Martelli, 2014) and some grape nursery were found to present leafroll symptoms, allowing the assumption of infection by GLRaV-2 which is found almost associated with GLRaV-3 and propagation by infected shoot.

Lekikot *et al.* (2012) reported also the presence of GLRaV-2 in Algeria with less prevalence. The fact of this large propagation of GLRaV-2 may be due to the use of infected plant propagating materials. Indeed, observations made on some grape nursery during the prospection reveals the presence of leafroll symptoms. (Lekikot, 2012) reported that native varieties were more infected than the imported ones which may be due to large movements of infected material. Further studies on grape nursery were needed in order to understand the propagation of this virus in Algeria. Few studies were dedicated to the genetic diversity of GLRaV-2. Based on CP gene analyses, (Bertazzon *et al.*, 2010) reported five clades. In the same year, (Jarugula *et al.*, 2010) reported six lineages.

In this study, phylogenetic analyses of HSP70h gene performed reveal the presence of six lineages confirming previous studies. Indeed, comparison at the nucleotide level reveals that the described groups present less than 86% similarity between them and the similarity within groups reveal more than 95% identity. Furthermore, similarity plot reveals five distinct variants except for variant PV20 due to the absence of full length sequence. Our results were in concordance with those obtained by (Jarugula *et al.*, 2010) which reported an interlineage sequence identities between PN, 93/955, and H4 lineages of 83 to 86% and between 'PV20', BD, and RG lineages were 68 to 80%.

Phylogenetic tree reveals that all Algerians isolates provided from a distant region belong to the Group PN containing in general American isolates. Tree reveals the presence of isolates provided from a distant region and cluster in the same clade. The same result was obtained by several authors who rejected the hypothesis that the phylogeny of GLRaV-2 population depend on geographical origin. Thus, (Jarugula *et al.*, 2010) reported that isolates provided from distant vineyards show an important similarity and cluster in the same lineage. This observation may be due to the large movement of variants among root stock and cultivars. Indeed, a large survey on autochthonous and wild populations is needed for a better understanding of genetic variation of a GLRaV-2 population. However Results show that the groups PN and H4 were the most important group constituted in the majority of North American isolates, this result may be due to the large sequences providing from this region.

The high genetic stability of viruses can be attributed to negative or purifying selection to maintain the functional integrity of the viral genome. However, several genera were reported as evolving under positive selection (the genera Aureusvirus, Carmovirus, Dianthovirus, Necrovirus and Tombuvirus). Thus (Jarugula *et al.*, 2010) reported that HSP70h evolve under negative selection with $dN/dS < 1$. In our study, we found that this gene evolve under positive selection. Indeed, we obtained the mean ratio dN/dS greater than 1 ($dN/dS = 1,81$ for SLAC and $dN/dS = 3,71$). Results reveal also that some sites were under neutral selection and others under negative selection. Indeed, some sites in the HSP70h were exposed to mutation which were maintained in the population and other were under stability. To understand whether codon may be predisposed to mutation. The positive selection obtained may be explained by the changes on amino acid in some site of HSP70h gene, indicating specificity of each variant. It is interesting to know the effect of these mutations on plant virus interaction. Indeed, these mutations may explain the differences in virulence between the different variants. (Bertazzon *et al.*, 2010) reported that isolates from group BD were less virulent and were unable to induce graft incompatibility and it rarely caused leafroll symptom. In contrast, the RG isolates appeared to be more virulent.

The absence of recombination events may be explained by the fact that each vine may be infected by only one

variant limiting genome exchange between distant variants. Indeed, results reveal the presence of only one variant in Algeria and the absence of natural vector of GLRaV-2 limit the large distribution of variants. Further studies of variant distribution are necessary.

REFERENCES

- Abou-Ghanem, N. and S. Sabanadzovic and A. Minafra and P. Saldarelli and G. Martelli. 1998. Some properties of Grapevine leafroll-associated virus 2 and molecular organization of the 3' region of the viral genome. *Journal of Plant Pathology*. 37-46.
- Angelini, E. and N. Bertazzon and M. Borgo. 2004. Diversity among Grapevine Leafroll-associated Virus 2 Isolates Detected by Heteroduplex Mobility Assay. *Journal of Phytopathology*. 152: 416-22.
- Bertazzon, N. and M. Borgo and S. Vanin and E. Angelini. 2010. Genetic variability and pathological properties of Grapevine leafroll-associated virus 2 isolates. *European Journal of Plant Pathology*. 127: 185-97.
- Beuve, M. and L. Sempé and O. Lemaire. 2007. A sensitive one-step real-time RT-PCR method for detecting Grapevine leafroll-associated virus 2 variants in grapevine. *Journal of Virological Methods*. 141: 117-24.
- Clark, M. F. and A. Adams. 1977. Characteristics of the microplate method of enzyme-linked immunosorbent assay for the detection of plant viruses. *Journal of general virology*. 34: 475-83.
- Ghanem-Sabanadzovic, N. A. and S. Sabanadzovic and M. Castellano and D. Boscia and G. Martelli. 2000. Properties of a new isolate of Grapevine leafroll-associated virus 2. *VITIS-GEILWEILERHOF-*. 39: 119-22.
- Goszczynski, D. and G. Kasdorf and G. Pietersen and H. Van Tonder. 1996. Detection of two strains of grapevine leafroll-associated virus 2. *VITIS-GEILWEILERHOF-*. 35: 133-6.
- Greif, C. and R. Garau and D. Boscia, *et al.* 1995. The relationship of grapevine leafroll-associated closterovirus 2 with a graft incompatibility condition of grapevines. *Phytopathologia Mediterranea*. 167-73.
- Gugerli, P. and J. Brugger and R. Bovey. 1984. L'enroulement de la vigne: mise en évidence de particules virales et développement d'une méthode immuno-enzymatique pour le diagnostic rapide. *Revue suisse de viticulture, arboriculture, horticulture*.
- Jarugula, S. and O. J. Alabi and R. R. Martin and R. A. Naidu. 2010. Genetic variability of natural populations of Grapevine leafroll-associated virus 2 in Pacific Northwest vineyards. *Phytopathology*. 100: 698-707.
- Karasev, A. V. 2000. Genetic diversity and evolution of closteroviruses. *Annual review of phytopathology*. 38: 293-324.
- Lekikot, A. K., Elbeaino, T., Ghezli, C., and Digiario, M. 2012. *Proceedings of the 17th Congress of the International Council for the Study of Preliminary Survey of Grapevine Viruses in Algeria, Davis, California, USA, October 7-14, 2012*. University of California, Foundation Plant Services.
- Martelli, G. and N. Abou Ghanem-Sabanadzovic and A. Agranovsky, *et al.* 2012. Taxonomic revision of the family Closteroviridae with special reference to the grapevine leafroll-associated members of the genus Ampelovirus and the putative species unassigned to the family. *Journal of Plant Pathology*. 94: 7-19.
- Martelli, G. P. 2014. Directory of virus and virus-like diseases of the grapevine and their agents. *Journal of Plant Pathology*. 96: S1-S136.
- Meng, B. and C. Li and D. E. Goszczynski and D. Gonsalves. 2005. Genome sequences and structures of two biologically distinct strains of Grapevine leafroll-associated virus 2 and sequence analysis. *Virus genes*. 31: 31-41.
- Moury, B. and V. Simon. 2011. dN/dS-based methods detect positive selection linked to trade-offs between different fitness traits in the coat protein of potato virus Y. *Molecular biology and evolution*. 28: 2707-17.
- Rowhani, A. and Y. Zhang and D. Golino and J. Uyemoto. 2000. Isolation and partial characterization of two new viruses from grapevine. *Proceedings of XIII International Council for the Study of Viruses and Virus-Like Diseases of the Grapevine, Adelaide*. 82.
- Rowhani, A. and Y. Zhang and D. Golino and J. Uyemoto. 2002. Isolation and characterization of a new closterovirus from grapevine. *Phytopathology*. 92: S71.
- Saitou, N. and M. Nei. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular biology and*

evolution. 4: 406-25.

Svanella-Dumas, L. and M. Dulucq and T. Candresse and P. Gentit and X. Foissac. Polyvalent detection of fruit tree tricho, capillo and foveaviruses by nested RT-PCR using degenerated and inosine containing primers (PDO RT-PCR). *Proceedings of the XVIII International Symposium on Virus and Virus-like Diseases of Temperate Fruit Crops-Top Fruit Diseases 550, 2000, 37-44.*

Tamura, K. and G. Stecher and D. Peterson and A. Filipksi and S. Kumar. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular biology and evolution.* 30: 2725-9.

Thompson, J. D. and D. G. Higgins and T. J. Gibson. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic acids research.* 22: 4673-80.

Zhu, H. and K. Ling and D. Goszczynski and J. Mcferson and D. Gonsalves. 1998. Nucleotide sequence and genome organization of grapevine leafroll-associated virus-2 are similar to beet yellows virus, the closterovirus type member. *Journal of general virology.* 79: 1289-98.